

# Opposing roles of the aldo-keto reductases AKR1B1 and AKR1B10 in colorectal cancer

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## Abstract

**Purpose** Aldo-keto reductases (including AKR1B1 and AKR1B10) constitute a family of oxidoreductases that have been implicated in the pathophysiology of diabetes and cancer, including colorectal cancer (CRC). Available data indicate that, despite their similarities in structure and enzymatic functions, their roles in CRC may be divergent. Here, we aimed to determine the expression and functional implications of AKR1B1 and AKR1B10 in CRC.

**Methods** AKR1B1 and AKR1B10 gene expression levels were analyzed using publicly available microarray data and *ex vivo* CRC-derived cDNA samples. Gene Set Enrichment Analysis (GSEA), The Cancer Genome Atlas (TCGA) RNA-seq data and The Cancer Proteome Atlas (TCPA) proteome data were analyzed to determine the effect of high and low AKR1B1 and AKR1B10 expression levels in CRC patients. Proliferation, cell cycle progression, cellular motility, adhesion and inflammation were determined in CRC-derived cell

lines in which these genes were either exogenously overexpressed or silenced.

**Results** We found that the expression of AKR1B1 was unaltered, whereas that of AKR1B10 was decreased in primary CRCs. GSEA revealed that, while high AKR1B1 expression was associated with increased cell cycle progression, cellular motility and inflammation, high AKR1B10 expression was associated with a weak inflammatory phenotype. Functional studies carried out in CRC-derived cell lines confirmed these data. Microarray data analysis indicated that high expression levels of AKR1B1 and AKR1B10 were significantly associated with shorter and longer disease-free survival rates, respectively. A combined gene expression signature of AKR1B10 (low) and AKR1B1 (high) showed a better prognostic stratification of CRC patients independent of confounding factors.

**Conclusions** Despite their similarities, the expression levels and functions of AKR1B1 and AKR1B10 are highly divergent in CRC, and they may have prognostic implications.

**Keywords** AKR1B1 · AKR1B10 · Colorectal cancer · Inflammation · Motility · Prognosis

Betul Taskoparan and Esin Gulce Seza share equal contribution.

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## 1 Introduction

Recent studies have indicated that the incidences of diabetes and cancer are closely related in many (but not all) countries worldwide [1, 2] and that glucose sensitizing drugs can provide significant protection from the development of colorectal cancer (CRC) [3]. Therefore, an evaluation of signaling pathways that may be affected in both diabetes and CRC is warranted. The aldo-keto reductases (AKRs) constitute a large family of oxidoreductases that can catalyze reduction reactions in the presence of nicotinamide adenine dinucleotide (phosphate) (NAD(P)H) [4]. AKRs, most commonly

AKR1B1, are involved in the first step of the polyol pathway, where conversion of glucose into sorbitol takes place using NAD(P)H as cofactor. As such, this enzyme has been found to be implicated in the pathophysiology of diabetes [5]. AKR1B10 only weakly reduces glucose, but has been found to be implicated in the metabolism of compounds such as 4-hydroxynonenal, acrolein, retinals and phospholipid aldehydes [6]. These compounds can also be reduced by AKR1B1 [6]. AKRs have also been found to be involved in the reduction of lipid peroxidation-derived aldehydes and their corresponding glutathione conjugates [7]. These reduced compounds serve as inflammatory signals, which mediate reactive oxygen species (ROS)-related signaling and lead to inflammatory responses [8]. Inflammation has been found to be strongly implicated in the development of CRC [9] and inhibition of the ubiquitously expressed AKR protein AKR1B1 with fidarestat has been found to inhibit inflammation in CRC [10]. Additionally, siRNA-mediated silencing of AKR1B1 in the colon cancer-derived cell line SW480 was found to inhibit tumor growth in a nude mouse xenograft model [11]. AKR1B1 inhibition with fidarestat has been shown to reduce the expression of the oncogenic microRNA miR-21, leading to upregulation of the tumor suppressor proteins PTEN [12] and PCDC4 [13] in CRC.

Data on the role of AKR1B10 in CRC is relatively limited. AKR1B10 is abundantly expressed in non-transformed small intestine and colon, while a lower expression has been observed in various other organs such as liver, prostate, thymus, testis and skeletal muscle [14]. A significant decrease in the expression of AKR1B10 has been reported for CRCs and adenomas compared to their corresponding normal tissues [15]. Additionally, AKR1B10 has been reported to serve as a direct transcriptional target of p53 and to participate in p53-mediated apoptosis [15]. AKR1B10 can also metabolize electrophilic carbonyl compounds to harmless intermediates and, thereby, protect CRC cells from DNA damage [16].

Thus, in spite of the high similarities in structures and enzymatic activities of AKR1B1 and AKR1B10, the expression levels and functions of the two proteins in CRC appear to be divergent. So far, however, a systematic functional comparison of the two proteins in CRC has not been carried out. Here, we have experimentally established a complete divergence in activation of inflammatory signaling and cellular motility of the two proteins. Gene set enrichment analysis (GSEA) using a publicly available microarray dataset as well as The Cancer Genome Atlas (TCGA) and The Cancer Proteome Atlas (TCPA) data analyses for tumors with high and low AKR1B1 expression levels or high and low AKR1B10 expression levels confirmed these findings. AKR1B1 expression was also found to be associated with oncogenic characteristics, since silencing of the corresponding gene led to reduced cell proliferation and a slower cell cycle progression. Additionally, multivariate Cox proportional hazards regression analysis revealed that a

combination of high AKR1B10 expression and low AKR1B1 expression was of prognostic significance, i.e., it was significantly associated with a longer disease-free survival of CRC patients, independent of confounding factors.

## 2 Materials and methods

### 2.1 Cell culture and transfection

HCT-116 cells were purchased from the Deutsche Sammlung von Mikroorganismen und Zellkulturen (DSMZ, Braunschweig, Germany) and HT-29 cells from ŞAP Enstitüsü, Ankara, Turkey. SW480, RKO, Caco-2 and LoVo cells were purchased from the ATCC (Middlesex, UK). All cell lines were cultured under ATCC-specified conditions in a humidified incubator with 5% CO<sub>2</sub> at 37 °C. All cell culture consumables were purchased from Biochrom AG, Germany. Normal colon RNA was purchased from Origene (Rockville, MD, USA).

AKR1B1 expression was knocked down in HCT-116 cells using a SureSilencing shRNA plasmid kit (Catalog no: KH02359, Qiagen, Germany). A pool of 4 shRNA vectors was transfected into ~70% confluent HCT-116 cells in OptiMEM (Thermo Fischer Scientific, USA) using X-tremegene HP at a 1:2 ratio (1 µg vector:2 µl of X-tremegene HP) according to the manufacturer's protocol. Transfected cells were selected using 500 µg/ml G418 (Roche, Switzerland) for 3–4 weeks after which two clonal lines with a stable knockdown of AKR1B1 were chosen for further experiments. As a control, cells stably transfected with a scrambled control vector as provided in the kit was used. The cells were maintained in 225 µg/ml G418. All experiments with stably transfected cells were carried out within the 8th passage. For exogenous overexpression of AKR1B1, a pCMV6-AC mammalian expression vector was used (Origene). Cells were transfected at 70% confluency using X-tremegene HP at 1:2 ratio of plasmid to transfection reagent. After 24 h the cells were harvested and processed for protein or RNA isolation. Where indicated, rescue experiments were carried out by overexpressing AKR1B1 in AKR1B1 silenced cells (shB1\_Clone 1).

AKR1B10 cDNA, cloned into a pCOLD1 vector, was obtained as a gift from Dr. Satoshi Endo, Gifu Pharmaceutical University, Japan [6]. The AKR1B10 cDNA insert was excised using *NheI* and *EcoRI* and sub-cloned into a pCDNA3.1 vector using the same restriction sites. Correct insertion was confirmed by sequencing. For exogenous overexpression, HCT-116 cells were transfected at 70% confluency using X-tremegene HP at 1:1 ratio of plasmid to transfection reagent. After 24 h the cells were harvested and processed for protein or RNA isolation. All transient overexpression and silencing experiments were confirmed by Western blotting.

## 2.2 Protein isolation and Western blotting

Total protein was isolated using a M-PER Mammalian Protein Extraction Reagent (Thermo Fisher Scientific, USA) containing protease and phosphatase inhibitors (Roche, Germany) according to the manufacturer's instructions. For the isolation of cytoplasmic and nuclear proteins cells were collected, washed with PBS and resuspended in hypotonic buffer (100 mM HEPES pH:7.5, 40 mM NaF, 100  $\mu$ M Na<sub>2</sub>MoO<sub>4</sub> and 1 mM EDTA). Next, the cells were transferred to pre-chilled eppendorf tubes and incubated on ice for 15 min. Then, 40  $\mu$ l of 10% NP-40 was added and mixed, after which the tubes were centrifuged. The supernatant was carefully collected as cytoplasmic fraction. Next, the pellets were resuspended in nuclear extraction buffer (20 mM HEPES pH:7.9, 0.2 mM EDTA, 3 mM MgCl<sub>2</sub>, 840 mM NaCl, 20% glycerol) by shaking for 30 min in an orbital shaker with vortexing every 15 min. The resulting suspension was centrifuged at 14000 $\times$ g for 10 min at 4 °C after which the supernatants were transferred to fresh eppendorf tubes as nuclear fraction. The amounts of proteins were measured using a Coomassie Protein Assay Reagent (Thermo Fisher Scientific, USA) and determined relative to a standard curve generated with bovine serum albumin. For subsequent Western blotting 30–50  $\mu$ g of proteins from each sample were separated in 10% SDS- polyacrylamide gels and transferred to polyvinylidene fluoride (PVDF) membranes. Western blotting was carried out using standard protocols. Visualization of the bands was performed using a Clarity ECL Substrate (Bio-Rad, USA) and imaged on a Chemi-Doc MP system (Bio-Rad). The membranes were also incubated with anti- $\beta$ -Actin or anti-GAPDH antibodies to ensure equal protein loading. Details on the antibodies used and their dilutions are provided in Supplementary Table 1.

## 2.3 cDNA synthesis and qRT-PCR

Total RNA was isolation using a RNeasy RNA Extraction Kit (Qiagen, Hilden, Germany) according to the manufacturer's guidelines. The RNA was treated with DNase I (Thermo Scientific) to remove genomic DNA, after which 1  $\mu$ g RNA was converted to cDNA using a RevertAid First Strand cDNA Synthesis Kit (Thermo Scientific). qRT-PCR reactions were carried out in a Rotor Gene Q 6000 system (Qiagen) according to the manufacturer's instructions. Fold changes were calculated relative to an internal control ( $\beta$ -Actin) using the Pfaffl method [17]. MIQE guidelines were followed in the qRT-PCR reactions [18]. The primer sequences are listed in Supplementary Table 1.

## 2.4 Proliferation assays

To determine the effect of AKR1B1 expression knockdown on cell proliferation a BrdU incorporation assay was used. To

this end, 10,000 cells were seeded per well in a 96-well plate and allowed to attach for 24 h. Next, the cells were incubated with serum-free medium overnight for synchronization [19], after which complete medium was added and at 0, 24 and 48 h the medium was removed followed by a BrdU assay according to the manufacturer's instructions (Roche). Measurements were performed using a microplate reader (Thermo Fisher Scientific) at 370 nm within 5–15 min. To determine the effect of exogenous AKR1B10 overexpression, a Trypan blue exclusion assay followed by cell counting was carried out. Twenty-four hours after transfection, the cells were collected and mixed with a 0.4% Trypan Blue solution (Bio Rad) after which the cells were counted in a TC20™ Automated Cell Counter (Bio Rad).

## 2.5 Cell cycle assay

To determine the effects of AKR1B1 expression knockdown or exogenous AKR1B10 overexpression on cell cycle progression, the cells were synchronized overnight in serum free medium. Next, at 0 and 6–8 h for AKR1B1 knockdown and 24 h for AKR1B10 overexpression, the cells were collected and fixed through a drop-wise addition of 70% ice-cold ethanol and kept at -20 °C for at least 2 h. Next, the fixed cells were washed in PBS, resuspended in a staining solution consisting of 0.1% Triton X-100, 2 mg/ml RNase A (DNase free) and 20  $\mu$ g/ml Propidium Iodide (Sigma-Aldrich, USA) and incubated for 30 min in the dark at room temperature. The percentages of cells in different stages of the cell cycle were determined using the FL-3 channel of an Accuri C6 Flow Cytometer (BD Biosciences, USA) and compared with scrambled or empty vector controls.

## 2.6 Luciferase assay

In order to determine the effects of AKR1B1 and AKR1B10 on the transcriptional activity of NF- $\kappa$ B, a Pathdetect reporter plasmid (Agilent Genomics, USA), which contains 5 copies of the binding sites for NF- $\kappa$ B upstream of a firefly gene, was used. A pRL-TK Renilla plasmid (Promega, USA) was used as an internal control. 50,000 cells per well were seeded and allowed to attach in 48 well-plates. Next, the cells were transfected with a 1:250 ratio of Firefly:Renilla vectors for 24 h, harvested and assayed using a Dual-Glo Luciferase Assay Kit (Promega, USA) according to manufacturer's guidelines in conjunction with the use of opaque 96-well plates in a luminometer (Turner Biosystems, USA).

## 2.7 Determination of ROS levels

To determine whether AKR1B1 or AKR1B10 expression modification alters the level of reactive oxygen species (ROS) production, a nitro-blue tetrazolium (NBT) assay

was carried out. NBT is a soluble nitro-substituted aromatic tetrazolium compound that forms formazan crystals in the presence of cellular superoxide ions that can subsequently be measured colorimetrically. HCT-116 cells with a stable knockdown of AKR1B1 expression were seeded in 96-well plates ( $1 \times 10^4$  cells/well). 24 h after seeding, the cells were incubated with NBT solution (2 mg/ml for each well) for 5 h at 37 °C. Then, the cells were fixed with 100% methanol for 5 min and air dried at room temperature. The blue formazan crystals that were formed were solubilized in 120  $\mu$ l KOH and 140  $\mu$ l DMSO by thorough pipetting. The absorbance was read colorimetrically at 620 nm in a microplate reader.

### 2.8 *In vitro* scratch wound healing assay

To determine the effect of AKR1B1 and AKR1B10 expression on cellular motility, a scratch wound healing assay was carried out as described previously [20]. Briefly, stably transfected cells were seeded to 80% confluency. Next, scratch wounds were made in the respective plates using a sterile 100  $\mu$ l-pipette tip. Cell debris was removed by washing the cells twice with cell culture grade PBS. Then, the cells were incubated in complete medium containing 0.5  $\mu$ M mitomycin C [21] to prevent any cell proliferation and at 0, 24 and 48 h time points, the cell images were captured using a JuLi Smart Fluorescent Cell Imager. Medium containing 0.5  $\mu$ M mitomycin C was refreshed after 24 h. The width of the wounds at each time point was measured using a ruler.

### 2.9 Transwell migration assay

In order to assess the effects of AKR1B1 or AKR1B10 expression on cellular migration, a Transwell migration assay was carried out. To this end, the cells were starved overnight, washed twice and resuspended in medium containing 1% FBS. Then, the cells were counted using a hemocytometer and  $\sim 5 \times 10^4$  cells in a 100  $\mu$ l suspension were seeded on Transwell chambers containing membranes with 8  $\mu$ m pores (ThinCert™ Cell Culture Inserts, Greiner Bio-One, Germany). At the same time, the lower chamber was filled with complete medium (10% FBS). The cells were allowed to migrate for 48 h after which the Transwells were taken out and non-migrated cells were removed with sterile cotton swabs. This latter step was repeated at least twice. Next, the Transwells were fixed in 100% methanol for 10 min, stained with Giemsa solution (Merck Millipore, USA) for 2 min at room temperature, and washed with sterile distilled water extensively in order to remove all excess dye. Finally, the Transwells were left to air dry inside a fume hood. When the Transwell filters were completely dried, the membranes were cut out with a bistoury and mounted on a glass slide with a drop of

immersion oil. The total number of cells was counted at 20 $\times$  magnification under an inverted light microscope (Leica, Germany).

### 2.10 Collagen deposition assay

To determine whether AKR1B1 expression knockdown alters cellular motility through collagen deposition, a Sirius red assay was carried out. Briefly, stably AKR1B1silenced HCT-116 cells ( $5 \times 10^5$ ) were seeded in 12-well plates. 24 h after seeding, the cells were washed once with cell culture grade PBS and then fixed in Bouin's fluid (prepared as mixture of 15 ml saturated picric acid, 5 ml 37% formaldehyde and 1 ml glacial acetic acid) for 1 h. Next, the wells were rinsed 3 $\times$  in PBS for 15 min, after which the plates were air-dried and stained with 0.5 ml of 100 mg/ml Sirius Red (Sigma-Aldrich) dissolved in saturated picric acid for 1 h with mild shaking. Subsequently, the dye was removed and the wells were rinsed with 0.01 M HCl to remove the excess dye. The deposited collagen, which stains red, was visualized under an inverted light microscope.

### 2.11 Hanging drop assay

A hanging drop assay was carried out to determine whether exogenous AKR1B1 overexpression results in alterations in cell-cell adhesion as described previously [20]. Briefly, HCT-116 cells were transfected with 1 or 2  $\mu$ g AKR1B1 expression vector or its corresponding empty vector for 24 h and collected. Drops of these cells (30  $\mu$ l,  $2 \times 10^6$  cells/ml) were pipetted on the inner surface of the lid of a sterile low attachment Petri dish after which the lid was placed on the petri dish and the now 'hanging drops' were incubated for 48 h in a cell culture incubator. Next, the drops were pipetted onto glass slides, covered with coverslips and imaged. At least 20–30 aggregates were photographed for each transfection.

### 2.12 *In silico* analyses

Gene expression data of tumor datasets were downloaded from Gene Expression Omnibus (GEO) (<http://www.ncbi.nlm.nih.gov/geo>) and RMA normalized using BRB-array tools (<https://brb.nci.nih.gov/BRB-ArrayTools/>). Clinical data related to the GSE39582 dataset ( $n = 585$ ) were obtained from Marisa et al. [22]. Log expression values of AKR1B1 and AK1B10 were determined and plotted. To determine correlations between AKR1B1 or AKR1B10 expression and that of pro-inflammatory genes, Level3 RNA-seq and RPPA (reverse phase protein array)-based expression data of 132 primary CRC tissues were downloaded from The Cancer Genome Atlas (TCGA) data portal (<cancergenome.nih.gov>). Linear correlation (Spearman) analyses between AKR1B1 and AKR1B10 expression and a set of pro-inflammatory genes were performed. Tumor samples



from TCGA were independently ranked according to RNAseq-based AKR1B1 and AKR1B10 expression levels and tumors with top 30% and bottom 30% expression were analysed for each gene. Differential protein expression and GSEA analysis between tumors within the top and bottom 30% were performed for both genes using the Broad Institute's desktop application (<http://software.broadinstitute.org/gsea/downloads.jsp>) according to the corresponding guidelines. Collapsing mode was used as maximum probe and Gene Ontology "c5 all" was used as genesets database.

### 2.13 Determination of AKR1B1 and AKR1B10 expression in primary CRC samples

Twenty six annotated first-strand cDNA samples (6-normal, 4-stage I, 8-stage II, 4-stage IIIB and 4-stage IV) from the Human Tissue Scan Colon Cancer Tissue qPCR Panel IV, HCRT304 (Origene, USA), were diluted 1:10 after which relative AKR1B1 and AKR1B10 mRNA levels were measured by qRT-PCR using a CFX Connect Real-Time PCR Detection System (BioRad, USA). Cycle threshold (Ct) values of individual genes were subtracted from Ct values for the reference gene  $\beta$ -Actin ( $\Delta$ Ct), and subsequently used to calculate fold changes in relative gene expression levels ( $2^{-\Delta\Delta$ CT) relative to one of the normal samples. The primer sequences used are listed in Supplementary Table 1.

### 2.14 Statistical analyses

All experiments were carried out as 2 or 3 independent biological replicates, each with at least 3 technical replicates. GraphPad Prism 6.1 (GraphPad Software Inc., USA) or SPSS Statistics v.19 (IBM, 2010, Chicago, IL, USA) were used for data analysis. One-way ANOVA, Student's *t* test or Mann Whitney U test were employed to determine significance. Kaplan-Meier plots and the log-rank test were used to compare the clinical outcomes among high-low expression groups. Univariate and multivariate Cox proportional hazards regression analysis was performed using SPSS Statistics v.19.  $p < 0.05$  was considered as statistically significant. Log-rank tests using all cut-off values (LRMC) for a given dataset were obtained using an in-house, R-based script [23]. Specific statistical analyses are further explained in the figure legends.

## 3 Results

### 3.1 AKR1B1 and AKR1B10 expression in primary colorectal cancer samples and cell lines

To determine the expression of AKR1B1 and AKR1B10 in primary colorectal cancer (CRC) samples, publicly available gene expression data (microarray-based dataset

GSE39582;  $n = 585$ ) were downloaded from GEO. We found that the expression of AKR1B10 was significantly reduced in the CRC samples compared to normal samples, irrespective of cancer stage (all cancer stages versus normal:  $p < 0.001$ ) (Fig. 1A). In contrast, we found that the expression of AKR1B1 showed no difference between the CRC (all stages) and normal samples ( $p > 0.05$ ). A significant decrease in the expression of AKR1B1 was, however, observed in Stage 0 (carcinoma in situ) samples compared to normal control samples ( $p < 0.001$ ). To confirm the microarray-based expression data, we decided to determine the AKR1B1 and AKR1B10 mRNA expression levels in an independent set of CRC samples using qRT-PCR analysis. In concordance with the microarray-based data, we observed a significant decrease in AKR1B10 expression in the CRC samples compared to its respective normal samples (Fig. 1B), especially at stages III and IV (Supplementary Fig. 1A), whereas no significant changes in AKR1B1 expression were observed between these samples (Fig. 1B) or stages (Supplementary Fig. 1B).

We next determined the expression of AKR1B1 and AKR1B10 in several CRC-derived cell lines. Compared to normal colon, the mRNA expression of AKR1B1 was found to be higher in HCT-116 and Caco-2 cells and undetectable in all other cell lines tested. On the other hand, we found that the expression of AKR1B10 was relatively low in all cell lines tested, except HT29 (Fig. 1C). Also at the protein level, we found that AKR1B1 was expressed in HCT-116 and Caco-2 cells. An additional band appeared at a slightly higher molecular weight in HT-29 cells. A protein BLAST of all AKR family members revealed that AKR1B1 exhibits a 55% similarity in amino acid sequence with AKR1CL2, a 68% similarity with AKR1B15 and a 71% similarity with AKR1B10 (E.G.S. and S.B., unpublished data). Since we did not detect any mRNA expression of AKR1B1 in these cells, we speculate that the band observed may correspond to another member of the AKR family that is also recognized by the antibody used. AKR1B10 was found to be expressed solely in HT-29 cells (Fig. 1C). Therefore, to determine whether the expression of AKR1B1 or AKR1B10 plays any role in CRC, we separately silenced AKR1B1 and exogenously overexpressed AKR1B10 in HCT-116 cells (Fig. 1D). As a control, AKR1B1 and AKR1B10 were exogenously overexpressed in LoVo cells, which do not express any detectable levels of either protein (Supplementary Fig. 1C).

### 3.2 AKR1B1, but not AKR1B10, affects cellular proliferation and cell cycle progression

Using a BrdU incorporation assay, we found that AKR1B1 silencing resulted in a decrease in proliferation of HCT-116 cells (Fig. 2A). This observation was supported by a reduced phosphorylation of ERK1/2, a mitogenic marker of the MAP

kinase cascade, in cells grown in complete medium (Fig. 2C, left panel). To assess the effect of AKR1B10 on cellular proliferation, a Trypan blue exclusion assay followed by automated counting was carried out. No differences in cell numbers were observed when AKR1B10 was exogenously overexpressed in HCT-116, SW480 or LoVo cells (Fig. 2B), along with no changes in ERK1/2 phosphorylation (Fig. 2C, right panel).

To determine effect of AKR1B1 silencing on cell cycle progression, PI staining followed by flow cytometry was used. By doing so, we found that AKR1B1 silencing did not lead to a cell cycle arrest. Rather, a slowing down of the cell cycle was observed whereby a greater proportion of AKR1B1 silenced cells was retained in the G1 phase and entered the S phase later than in the scrambled control cells at 8 h (Fig. 2D, Supplementary Fig. 2A). Interestingly, we observed increased Cyclin E protein levels in starved (synchronized) and released (6–8 h) cells, as well as in unsynchronized cells in which AKR1B1 was silenced. However, when we determined the fold changes in Cyclin E levels of starved versus released cells for both AKR1B1 silenced and scrambled control cells, we observed a statistically significant decrease in Cyclin E in the AKR1B1 silenced cells 6–8 h after release. No difference was observed in Rb phosphorylation. Also, no difference in cell cycle progression was observed in exogenously AKR1B10 overexpressing HCT-116 cells (Supplementary Fig. 2B).

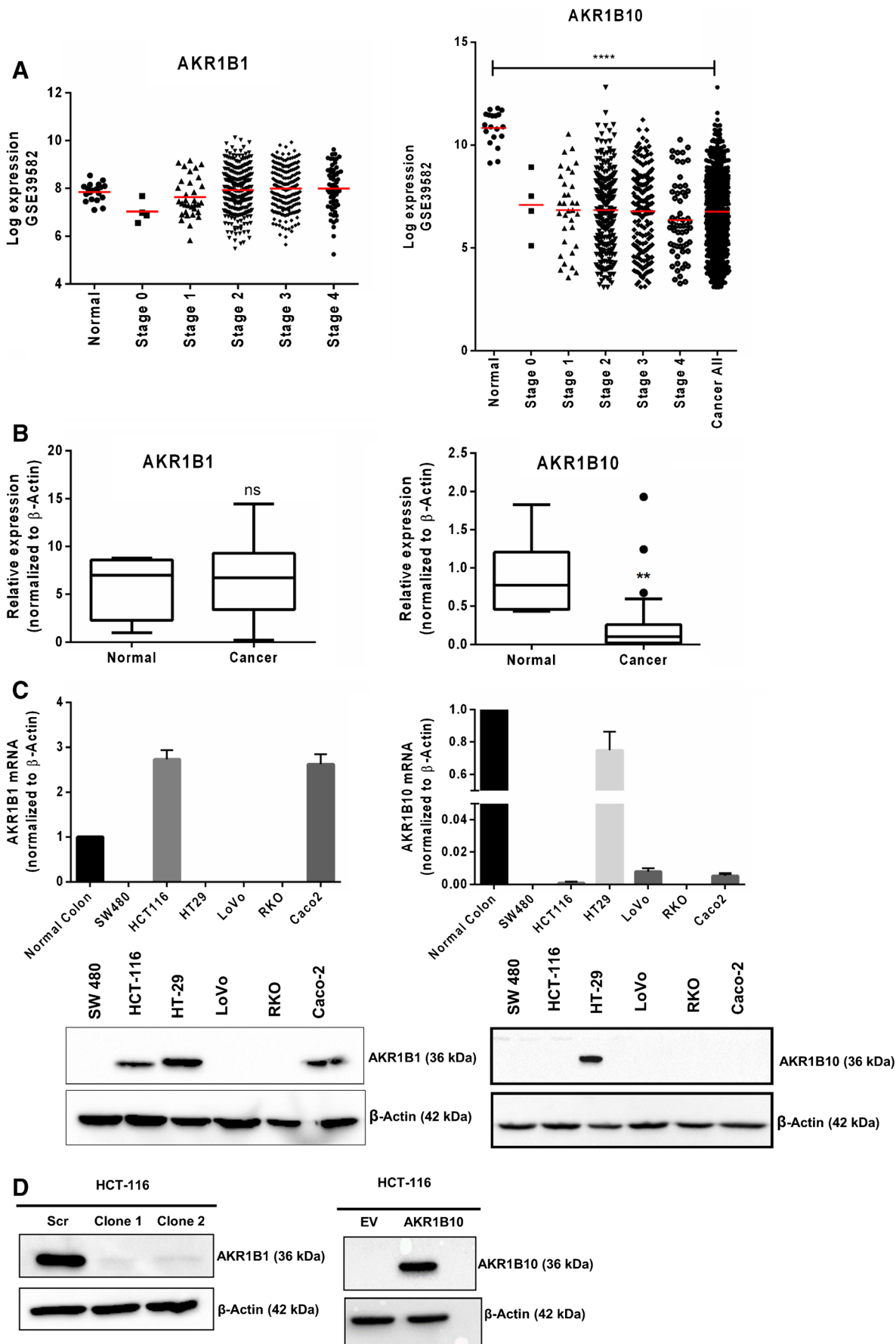
### 3.3 AKR1B1 enhances and AKR1B10 reduces cell motility

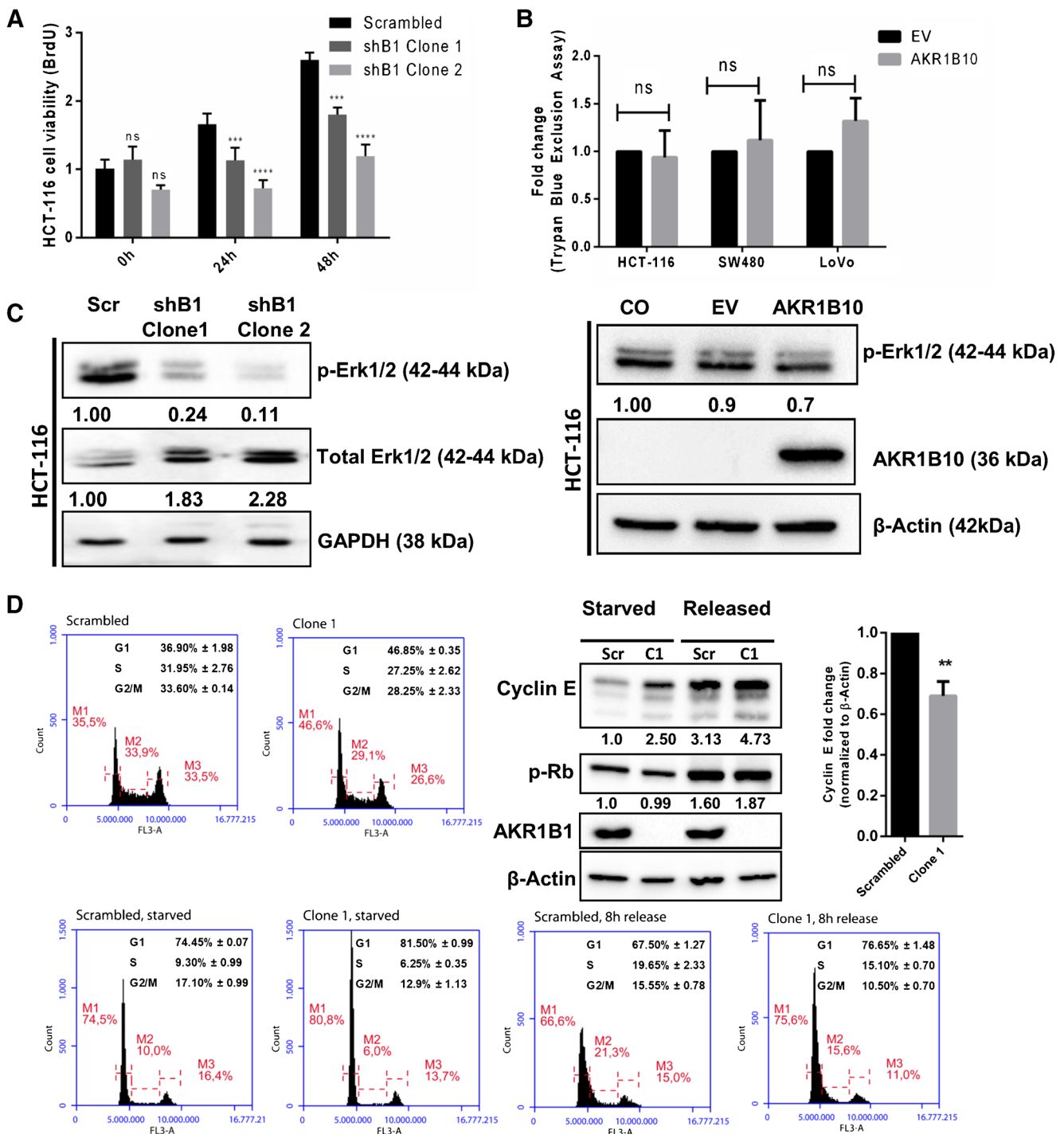
Gene Set Enrichment Analysis (GSEA) [24] of the GSE39582 [22] dataset for the high 30% and low 30% AKR1B1 expressing cases indicated a significant ( $p < 0.001$ ) enrichment in the Gene Ontology (GO) term “CELL\_SUBSTRATE\_JUNCTION” in tumor samples with a high AKR1B1 expression (Fig. 3A; for the full set of genes for this GO term, see Supplementary Table 2). The Cancer Proteome Atlas (TCPA) was used to mine data from colon and rectal patient samples [25]. Differential expression analysis of proteins between the AKR1B1 high 30% versus the AKR1B1 low 30% patients revealed a significantly higher expression of the mesenchymal/motility markers collagenVI ( $p = 0.0003$ ), fibronectin ( $p = 0.0002$ ) and transglutaminase ( $p = 0.0019$ ), whereas significantly lower expression levels were seen for the junctional protein claudin-7 ( $p = 0.0011$ ), which has been associated with epithelial characteristics [26], and FOXO 3a ( $p = 0.005$ ), a transcription factor that can enhance the expression of antioxidant proteins [27] (Fig. 3B; for the full list of significantly altered proteins see Supplementary Table 3). None of these proteins was found to be significantly up-regulated in AKR1B10 high expressing tumors ( $p > 0.05$ ).

Therefore, we set out to test whether silencing of AKR1B1 or exogenous overexpression of AKR1B10 may result in alterations in cell motility. We found that AKR1B1 silenced cells (both clones tested) showed a significantly slower closure in a wound healing assay (Fig. 3C and Supplementary Fig. 3A), while a Transwell migration assay revealed a significantly lower amount of migrating cells (Fig. 3D, left panel, Supplementary Fig. 3B). As the exogenous overexpression of AKR1B10 was carried out transiently, and we thus could not be sure that the cells at the border of the wound in the scratch wound assay would be expressing AKR1B10, we carried out a Transwell migration assay and found a significant decrease in cell migration when AKR1B10 was overexpressed (Fig. 3D, right panel, Supplementary Fig. 3C).

CollagenVI was found to be highly expressed ( $p = 0.0003$ ) in the TCPA data set of samples with high AKR1B1 expression. Therefore, we carried out a collagen deposition assay using Sirius Red staining on HCT-116 cells in which AKR1B1 was knocked down. We observed a decrease in collagen deposition in cells in which AKR1B1 was stably silenced (Fig. 3E). To further substantiate the notion that alterations in cell-cell adhesion in cells expressing AKR1B1 may lead to altered motility, we exogenously overexpressed AKR1B1 in HCT-116 cells using increasing amounts of plasmids and carried out a hanging drop assay. By doing so we found that, while control cells formed large cell clusters indicating good cell-cell contacts, AKR1B1 overexpressing cells showed a clear decrease in the formation of compact cellular aggregates (Fig. 3F). This effect was more pronounced in cells that were transfected with the higher amount of plasmid. Interestingly, we did not observe any change in expression of well-known epithelial markers, such as E-cadherin, or mesenchymal markers, such as vimentin (data not shown).

**Fig. 1 Expression of AKR1B1 and AKR1B10 in primary CRC samples and CRC-derived cell lines.** (A). Log<sub>2</sub> AKR1B1 and AKR1B10 expression values from the GSE39582 dataset ( $n = 585$ ) are plotted. The horizontal lines indicate means. Statistical significance was determined by Student's t-test. No change in AKR1B1 expression and a significant decrease in AKR1B10 expression was observed across stages I-IV compared to normal samples. (B). qRT-PCR analysis of CRC cDNA samples from a Tissue Scan array showing no difference in AKR1B1 expression and a reduction in AKR1B10 expression in CRC versus normal tissues. Statistical analysis was carried out using a non-parametric Mann Whitney U test. (C). qRT-PCR and Western blot-based AKR1B1 and AKR1B10 expression analyses of a panel of CRC-derived cell lines. (D). Western blot confirming the silencing of AKR1B1 (shB1 Clone 1 and shB1 Clone 2) and exogenous overexpression of AKR1B10 in HCT-116 cells. \*\*\*\* $p < 0.0001$ , \*\* $p < 0.01$





**Fig. 2 Effect of AKR1B1 and AKR1B10 expression on cellular proliferation and cell cycle progression.** (A). BrdU incorporation assay showing a decrease in proliferation in stably AKR1B1 silenced HCT-116 cells. Statistical analyses were carried out with ANOVA using Tukey's posthoc test. (B). Trypan blue exclusion assay showing absence of change in proliferation in HCT-116, SW-480 and LoVo cells transiently transfected with an AKR1B10 expression plasmid. (C). Western blot showing that AKR1B1 silencing leads to a decrease in EKR1/2 protein activation, whereas exogenous AKR1B10 overexpression did not result in any alteration. Numbers under the bands indicate band intensities normalized to the loading control. (D). Cell cycle distribution of serum

starvation-synchronized AKR1B1 silenced cells collected at 0 and 8 h after release from starvation. The average cell cycle distributions (three independent experiments) of scrambled control and shB1 Clone1 (unsynchronized or synchronized and harvested 6–8 h after release) cells are shown above representative histograms. The delay in cell cycle progression seen in these cells was corroborated by significantly lower fold changes in Cyclin E protein in shB1 Clone-1 cells before and after starvation compared to control cells. No difference in Rb phosphorylation was observed. Scr refers to control cells transfected with the scrambled plasmid, C1 refers to shB1 Clone 1, C2 refers to shB1 Clone 2, CO refers to cells only



### 3.4 AKR1B1 expression enhances and AKR1B10 inhibits NF- $\kappa$ B activity

Previously, tumors have been described as unhealed wounds that recruit a vast number of inflammatory cells that can provide a microenvironment that is conducive for further proliferation and metastasis [28]. To assess whether the altered motilities observed in the presence of AKR1B1 or AKR1B10 expression are also accompanied by alterations in inflammation, we performed a GSEA for CRC samples exhibiting the highest and lowest 30% AKR1B1 expression levels, using the GSE39582 dataset [22]. A highly significant ( $p < 0.001$ ) enrichment was observed for the GO term “REGULATION\_OF\_CYTOKINE\_PRODUCTION” in tumors with a high AKR1B1 expression (Fig. 4A; for the full set of genes for this GO term, see Supplementary Table 2). On the other hand, we found that samples exhibiting the highest 30% AKR1B10 expression did not show a significant enrichment of any GO terms related to inflammation.

Using RNA-seq data from the TCGA portal for primary CRC samples from 132 patients, we next carried out a Spearman correlation analysis between the expression of AKR1B1 or AKR1B10 and a set of 16 pro-inflammatory genes. We found that AKR1B1 expression was positively correlated with 10 of the 16 genes, whereas the expression of AKR1B10 was positively correlated with only 3 of the genes (Fig. 4B). Strikingly, no common gene could be identified that was significantly correlated with both AKR1B1 and AKR1B10.

In order to subsequently determine whether inflammatory signaling was altered in the cellular models used in the current study, we examined the activation of nuclear factor kappa B (NF- $\kappa$ B). NF- $\kappa$ B is a transcription factor that enhances the expression of pro-inflammatory genes such as cytokines, chemokines, adhesion molecules and other immunoregulatory mediators [29]. Nuclear translocation of NF- $\kappa$ B is associated with its transcriptional activation [30]. Nuclear fractions of cell lysates from stably AKR1B1 silenced HCT-116 cells or exogenously AKR1B10 overexpressing HCT-116 cells showed a reduced translocation of the NF- $\kappa$ B subunits p65 and p50 from the cytoplasm to the nucleus (Fig. 4C). Moreover, when we overexpressed AKR1B1 in AKR1B1-silenced HCT-116 cells, we observed a partial rescue in the nuclear translocation of p65 and p50 (Supplementary Fig. 4A). Additionally, we found a reduced NF- $\kappa$ B activation in luciferase reporter assays when AKR1B1 was silenced in HCT-116 cells. Exogenous overexpression of AKR1B1 in LoVo cells reversed this effect (Fig. 4D). On the contrary, we found that exogenous overexpression of AKR1B10 resulted in a reduced transcriptional activity of NF- $\kappa$ B in HCT-116 and LoVo cells (Fig. 4E). To further comprehend the mechanism underlying enhanced NF- $\kappa$ B activation, we

examined ROS production in the context of AKR1B1 or AKR1B10 expression. We found that AKR1B1 silencing resulted in a significantly lower amount of ROS formation (Supplementary Fig. 4B), whereas no change in ROS production was observed when AKR1B10 was exogenously overexpressed, indicating that for this other mechanisms must be at work (data not shown).

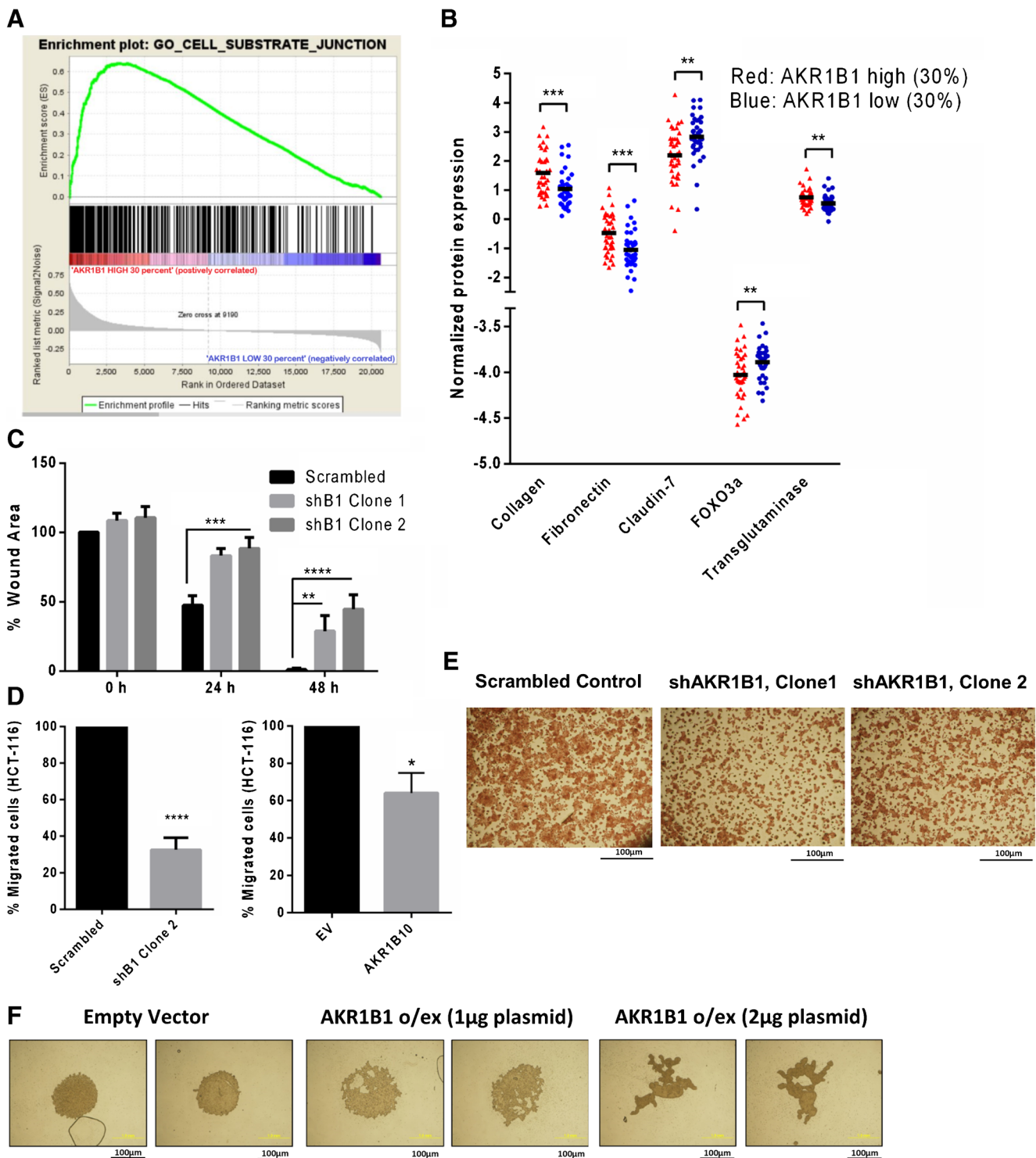
### 3.5 High AKR1B1 expression correlates with a poor prognosis and high AKR1B10 expression correlates with a good prognosis in CRC patients

We next queried whether the expression of AKR1B1 and AKR1B10 had any prognostic significance by analyzing the GSE39582 dataset [22]. Log-Rank tests were performed between two groups of patients stratified by all possible threshold values for each gene. The cut-off values that were within the 25–75 percentiles with the lowest  $p$  values were selected for further Kaplan-Meier analyses. By doing so, we found that a high AKR1B1 expression was associated with a shorter disease-free survival (DFS), while a high AKR1B10 expression was significantly associated with a longer DFS (Fig. 5A and B). Although we did not observe any statistically significant difference in AKR1B1 expression across different CRC stages in the GSE36582 set, the same dataset showed that a high AKR1B1 expression was associated with a shorter DFS in stage 2, 3 and 4 CRC (Supplementary Fig. 5).

To determine whether AKR1B1 and/or AKR1B10 may serve as prognostic indicators for overall CRC survival, we performed an independent multivariate Cox proportional hazards regression analysis (MVA), which revealed that this was indeed the case (Fig. 5C). Based on these observations, we generated a combined AKR gene signature for prognosis prediction, where AKR1B10-high and AKR1B1-low patients were classified as those with a “good” prognosis, AKR1B1-high and AKR1B10-low patients as those with a “bad” prognosis, and the rest as those with an “intermediate” prognosis (Fig. 5D). This classification generated a highly significant prognostic stratification with a log rank  $p$  value  $< 0.001$  (Fig. 5D). An additional MVA that included clinicopathological parameters revealed that the combined AKR gene signature was associated with disease-free survival independent of age, gender, *KRAS* or *BRAF* mutations, and TNM stage (Supplementary Table 4).

## 4 Discussion

The aldo-keto reductases AKR1B1 and AKR1B10 exhibit a high degree of similarity in amino acid sequence and structure and are able to reduce a number of common substrates [31]. However, AKR1B1 is ubiquitously expressed while the expression of



AKR1B10 is restricted to the gut and adrenal glands. Therefore, based on their distinct expression patterns, we hypothesized that AKR1B1 and AKR1B10 may play distinct roles in CRC. Here, we aimed to systematically test this hypothesis.

To this end, we first examined the expression of AKR1B1 and AKR1B10 in CRC tissues. Microarray and qRT-PCR-based data indicated that the expression of AKR1B1 was

similar in CRC and normal tissues whereas the expression of AKR1B10 was significantly reduced in the CRC samples, particularly in Stages III and IV. AKR1B10 has been reported to be a downstream target of p53. Therefore, loss of p53 activity may be implicated in loss of AKR1B10 expression [15]. However, p53 inactivation is not a universal phenomenon in CRC. Downregulation of AKR1B10 through promoter

◀ **Fig. 3 Effect of AKR1B1 and AKR1B10 expression on the metastatic behavior of CRC cells.** (A). GSEA showing a high enrichment score (ES) of the Gene Ontology term CELL\_SUBSTRATE\_JUNCTION in patients from the GSE39582 data set exhibiting the highest and lowest 30% AKR1B1 expression levels. The plot reflects the degree to which a gene set is overrepresented at the top or bottom of a ranked list of genes. The score at the peak of the plot (the score farthest from 0.0) is the ES for the gene set. The position of individual members of the gene set in the ranked list is indicated by vertical lines. (B). TCGA data showing significantly higher expression levels of the mesenchymal/motility markers collagen VI ( $p = 0.0003$ ), fibronectin ( $p = 0.0002$ ), and transglutaminase ( $p = 0.0019$ ) and a low expression of claudin-7 ( $p = 0.0011$ ) and FOXO 3a ( $p = 0.005$ ) in the high 30% AKR1B1 expressing samples versus the low 30% expressing samples. (C). Wound healing assay showing a significantly slower wound closure indicating a lower motility in stably AKR1B1 silenced HCT-116 cells. Statistical analyses were carried out using ANOVA and Tukey's posthoc test,  $*p < 0.05$ ,  $**p < 0.01$ . The cells were treated with mitomycin C (0.5  $\mu$ M) during the experiment to inhibit cell proliferation. (D). Transwell migration assay showing a slower migration of HCT-116 cells in which AKR1B1 was either silenced or AKR1B10 was overexpressed. Statistical analyses were carried out using Student's t-test,  $*p < 0.05$ ,  $***p < 0.0001$ . (E). Sirius Red assay showing less collagen deposition (less red color) in AKR1B1 silenced in HCT-116 cells. (F). Hanging drop assay showing a weaker cell-cell adhesion (less tight and well-formed aggregates) in a dose-dependent manner when HCT-116 cells were transfected with increasing amounts of the AKR1B1 overexpression (o/ex) plasmid

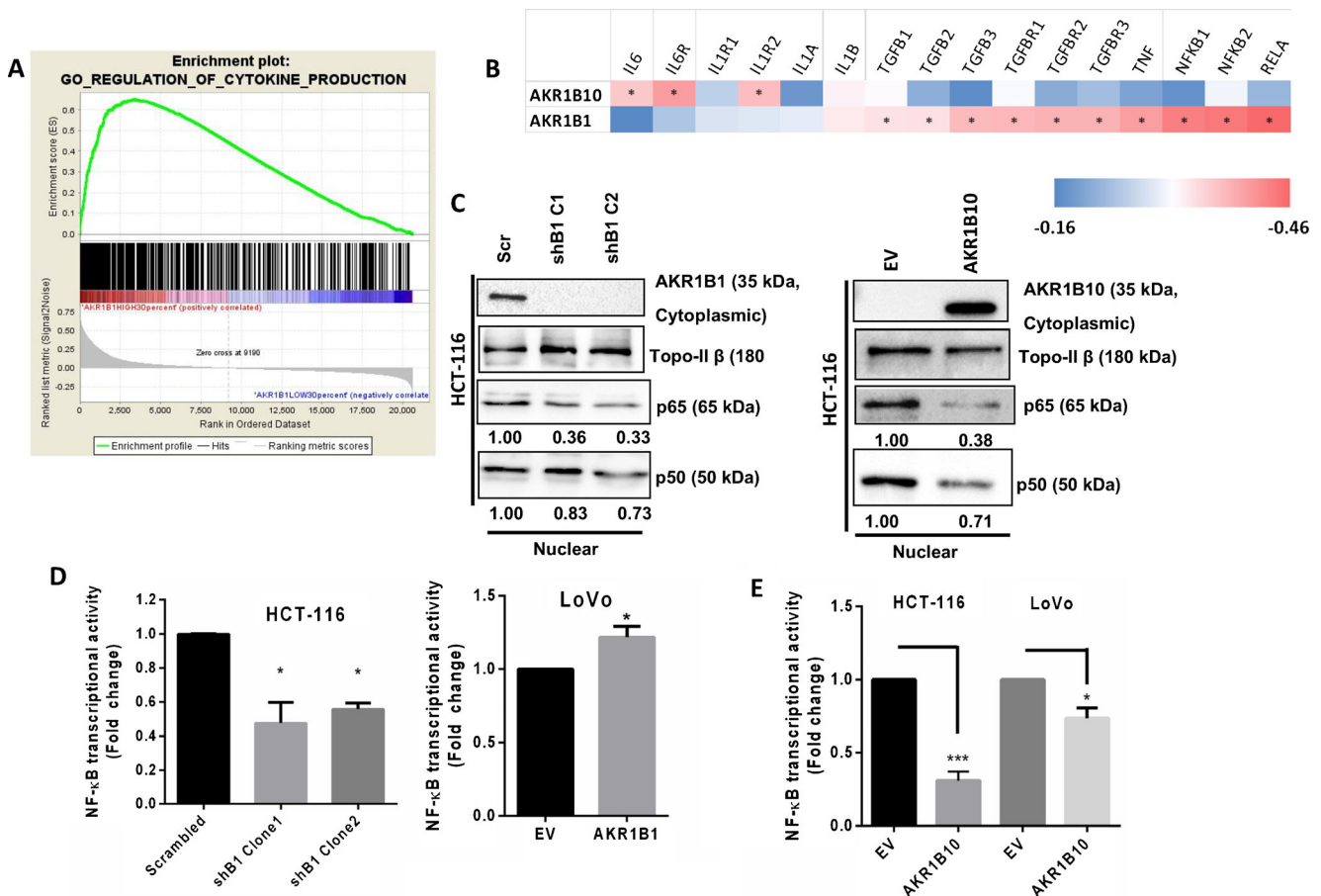
hypermethylation can also be ruled out since no CpG islands could be identified in the AKR1B10 promoter [15]. Moreover, using a cohort of 295 CRC samples present in the TCGA data set for which both methylation and expression information was available, we found that DNA methylation and RNA expression of AKR1B1 were significantly negatively correlated (rank of 143 among 201 genes with Bonferroni corrected  $p$  value  $< 0.01$ ,  $r$  value  $< -0.70$ ) while AKR1B10 was not found to be significantly correlated (S.D., A.O.G., S.B., unpublished data). Others have reported that in a cohort of 502 CRC cases, 70% of the tumors showed AKR1B1 promoter hypermethylation with similar levels of hypermethylation in both adenomas and carcinomas [32]. However, these authors also reported that re-expression of AKR1B1 was not observed in a DNMT (DNA Methyl Transferase) null CRC-derived cell line, thereby underscoring the complex regulation of AKR1B1 expression in CRC [32]. Hypermethylation of the AKR1B1 promoter is not restricted to CRC and has also been reported in e.g. breast cancer [33].

We found that AKR1B1 silenced cells showed a significantly reduced proliferation, without affecting apoptosis. We also observed downregulation of the mitogenic ERK1/2 proteins [34] in AKR1B1 silenced cells, which may underlie the reduced proliferation rate. Additionally, we found that a slowed down cell cycle may have contributed to the reduced proliferation rate of AKR1B1 silenced cells. Treatment of growth factor stimulated CRC-derived cells with the aldo-keto reductase inhibitor sorbinil has been shown to induce G1 arrest through downregulation of G1 cyclins and a reduced

activity of the transcription factor E2F [35]. In the current study, we found that 6–8 h after release from starvation-induced synchronization, a significantly larger proportion of AKR1B1 silenced cells was still in the G1 phase, whereas non-silenced control cells had already moved to the S phase. We also found that the control cells exhibited increased Cyclin E levels 6–8 h after release from starvation, as would be expected for cycling cells [36]. The increase in Cyclin E levels in the AKR1B1 silenced cells 6–8 h after release from starvation was significantly lower. This increase most likely reflects the greater accumulation of these cells in the G1 phase. Of note, we found that AKR1B1 silenced cells in general expressed higher levels of Cyclin E, irrespective of whether the cells were synchronized, starved or not. This was unlikely due to transcriptional upregulation as we did not observe any changes in the phosphorylation of Rb. Future studies are required to elucidate the mechanism underlying the high Cyclin E protein levels in AKR1B1 silenced cells. Interestingly, we found that alterations in chromosome segregation and cell cycle regulation were among the gene ontology terms that were enriched (albeit non significantly,  $p = 0.057$ ) in the GSEA of tumors that expressed low amounts of AKR1B1. Overexpression of AKR1B10 in HCT-116 cells did not result in any alteration in proliferation, apoptosis or cell cycle progression. AKR1B10 has been reported to metabolize cytotoxic carbonyl compounds to harmless intermediates [16]. Loss of AKR1B10 expression in CRC tissues may hamper the protection of these tissues from DNA damage by carbonyl compounds. It remains to be examined whether AKR1B10 overexpression can lead to alterations in cell cycle arrest in CRC cells with DNA damage.

Pharmaceutical inhibition of AKR1B1 in HT-29 cells has been shown to reduce growth factor-mediated adhesion to endothelial cells and expression of cell adhesion molecules [37]. These data were further supported by a reduced hepatic metastasis of KM20 cells in nude mice to which AKR inhibitors were administered [37]. Additionally, it has recently been found that AKR1B1 may serve as a target of the mesenchymal transcription factor Twist2 and as a major inducer of epithelial to mesenchymal transition in basal-like breast carcinomas [38]. Our analysis of CRC microarray-based expression data (GSEA), TCGA as well as functional studies on CRC-derived cell lines support the notion that high AKR1B1 levels may result in enhanced motility and migration. Conversely, we found that exogenous overexpression of AKR1B10 in HCT-116 cells resulted in a significantly slower cell motility. The role of AKR1B10 in metastasis appears to be highly context dependent. In breast cancer, AKR1B10 has been reported to be significantly associated with metastasis [39] through up-regulation of integrin  $\alpha 5$  and  $\delta$ -catenin [40] whereas, similar to our data, AKR1B10 overexpression in a nasopharyngeal carcinoma-derived cell line was found to result in a slower proliferation and a slower migration [41]. These differences may, at least partly, be explained by the different cell types





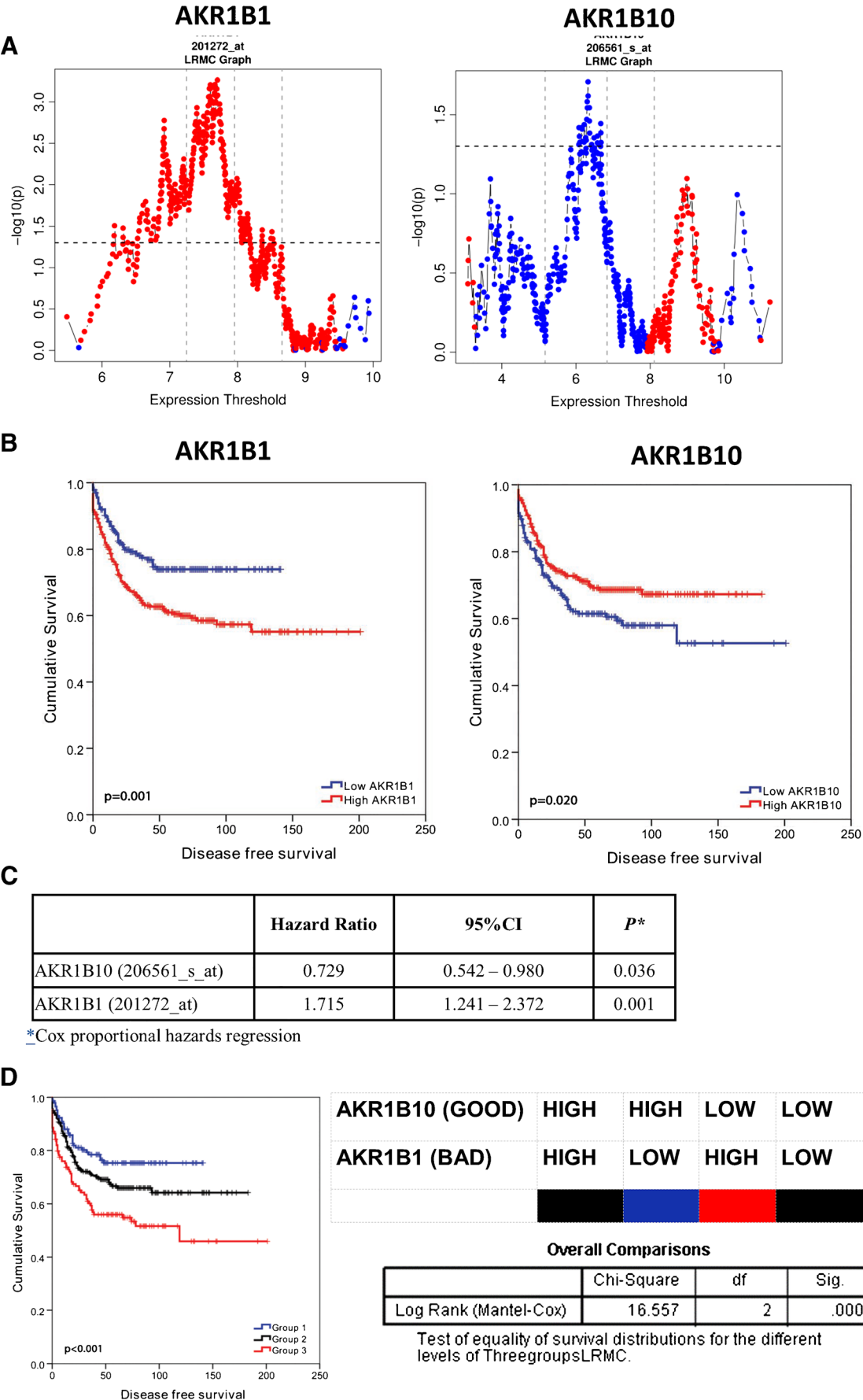
**Fig. 4 Effect of AKR1B1 and AKR1B10 expression of inflammation.** (A). GSEA showing high enrichment scores (ES) of the Gene Ontology term REGULATION\_OF\_CYTOKINE\_PRODUCTION in cases from the GSE39582 data set with the highest 30% AKR1B1 expression compared to the lowest 30%. The score at the peak of the plot (the score farthest from 0.0) is the ES for the gene set. The position of individual members of the gene set in the ranked list is indicated by vertical lines. (B). Using the CoadRead TCGA RNA-sequencing data, we tested whether 16 genes associated with inflammation correlated with either AKR1B1 or AKR1B10 in the same patients. A non-overlapping correlation of AKR1B1 and AKR1B10 was observed for several pro-inflammatory markers. Asterisks represent samples

that are significantly correlated. Correlation  $r$  values were marked from lowest (blue) to highest (red). (C). Western blot showing reduced nuclear translocation of NF- $\kappa$ B in stably AKR1B1 silenced and AKR1B10 overexpressing HCT-116. TopoII- $\beta$  was used as a loading control. Numbers under the bands indicate band intensities normalized to the loading control. (D). Luciferase assay showing reduced NF- $\kappa$ B transcriptional activity in stably AKR1B1 silenced HCT-116 cells. Overexpression of AKR1B1 in LoVo cells resulted in enhanced transcriptional activity. Exogenous AKR1B10 overexpression in both HCT-116 and LoVo cells resulted in reduced NF- $\kappa$ B transcriptional activity. Statistical analyses were carried out using Students  $t$ -test. \* $p < 0.05$ , \*\*\* $p < 0.001$

examined. The CRC-derived cell line HCT-8 has been found to transition from an epithelial (E) phenotype to a rare more rounded (R) highly metastatic phenotype when grown (20 days) on soft substrates [42]. The 'R' cells expressed significantly higher amounts of AKR1B10 than the 'E' cells. Thus, AKR1B10 expression may generally be reduced in epithelial type CRC cells that predominate in most of the models used to date. Interestingly, it has been found that epithelial type MCF-7 breast cancer-derived cells overexpressing AKR1B10 did not metastasize to lungs *in vivo*, whereas MDA-MB-231 cells that are more mesenchymal in nature [43] metastasized to the lungs when overexpressing AKR1B10 [40]. It remains to be seen whether primary CRC stem cells express higher amounts of AKR1B10 and whether this influences the metastatic capability of these cells.

**Fig. 5 AKR1B1 and AKR1B10 expression as prognostic CRC markers.** (A). LRMC plots for AKR1B1 and AKR1B10 showing  $p$  values from Log-Rank tests performed between these two groups stratified at each expression threshold value. Blue and red colors were used to indicate association of gene expression with a good or poor prognosis, respectively. The horizontal dotted line corresponds to the significance threshold of 0.05. Vertical lines correspond to expression thresholds of the 25th, 50th and 75th percentiles of patients. (B). Kaplan-Meier plots using patient data from the GSE39582 set based on cut-off levels within the 25-75th percentiles with the lowest  $p$  value for either AKR1B1 and AKR1B10 showed that high expression was significantly associated with shorter and longer disease-free survival rates for AKR1B1 and AKR1B10, respectively. (C). Multivariate Cox proportional hazards regression analyses with cut-offs in the 25-75th percentiles with the lowest  $p$  value were used for both genes, showing that the prognostic associations of AKR1B1 and AKR1B10 were independent of each other. (D). An AKR1B1 and AKR1B10 combined gene signature (AKR) can predict the prognosis of patients in the GSE39582 data set





Activation of the polyol pathway can lead to the development of oxidative stress which, in turn, can enhance the activation of inflammatory transcription factors such as NF- $\kappa$ B and AP-1 [8]. Pharmacological inhibition of AKR1B1 has been shown to reduce NF- $\kappa$ B activity in growth factor stimulated Caco-2 cells [11]. In support of these data, we found that exogenous AKR1B1 overexpression or silencing in CRC-derived cells was directly correlated with NF- $\kappa$ B activation, most likely through enhanced ROS production. No additional stimulation with growth factors was necessary to observe these effects. On the contrary, we found that exogenous AKR1B10 overexpression led to a reduced activation of NF- $\kappa$ B. According to TCGA CoadRead RNA-seq data, the expression of a number of distinct non-overlapping sets of inflammatory genes correlated with the expression of AKR1B1 or AKR1B10. We found that AKR1B1 was positively correlated with several pro-inflammatory genes. Among the genes that were positively correlated with the expression of AKR1B10, the highest positive correlation was observed for IL1-R2, a negative regulator of interleukin (IL)-1 that binds with high affinity to IL-1 $\beta$  and IL-1 $\alpha$ , but does not induce any downstream signaling [44]. Interestingly, the expression of IL-1 $\alpha$  was also found to be significantly positively correlated with AKR1B10, indicating the possibility of negative feedback mechanisms. While it is premature to conclude that AKR1B10 has anti-inflammatory properties, it is likely that the types of inflammation induced by AKR1B1 and AKR1B10 serve different purposes.

The prognostic significances of AKR1B1 and AKR1B10 individually have been reported for several different tumor types [15, 41, 45–47]. In CRC we found that the expression of AKR1B1 was not altered in cancer cells compared to normal cells, both *in silico* and in *in vitro* experiments. This observation is supported by another study in which no significant difference was observed in AKR1B1 expression in CRC, normal colon and ulcerative colitis tissue samples [48]. On the other hand, current and other studies [8, 10, 35, 49] indicate that AKR1B1 overexpression may be associated with alterations in proliferation, cell cycle progression and the activation of inflammatory pathways. AKR1B1 may also serve as a prognostic CRC marker since patients with a high AKR1B1 expression in the GSE39582 dataset showed a significantly shorter disease-free survival (DFS). We found that this notion was also supported by a lack of significant differences in the expression of AKR1B1 using qRT-PCR (TissueScan CRC patient cDNA array) on the basis of available TNM staging, node positivity and differentiation information (data not shown).

AKR1B10 expression is dependent on the site of tumor origin, with a generally lower expression reported for tumors of the head and neck, bladder, stomach and colon compared to their normal counterparts [50]. Moreover, in hepatocellular

carcinoma (HCC), where AKR1B10 has been reported to be strongly upregulated in Hepatitis B virus (HBV)-related tumors [51] as well as in primary hepatocellular carcinomas [46], AKR1B10 overexpressing patients have been found to have a more favorable prognosis, a reduced tumor recurrence and a longer survival [50, 51]. For CRC, a similar favorable prognosis has been noted for AKR1B10 high expressing tumors using three independent data sets [15]. We found that in the AKR1B10 high expressing group in the GSE39582 data set the corresponding patients also showed significantly longer disease-free survival rates. Importantly, we have shown here for the first time that a combined signature of low AKR1B1 and high AKR1B10 expression provides a better prognostic stratification for CRC patients compared to either gene alone, and that this stratification is independent of other confounding factors such as age, TNM stage and *KRAS* or *BRAF* gene mutations.

## 5 Conclusions

In the current study we have shown that the functional effects of alterations in AKR1B1 and AKR1B10 expression in CRC are highly divergent even though both proteins catalyze very similar reactions. Opposite effects were observed on cellular proliferation, cell cycle progression, cellular motility and activation of inflammatory signaling pathways. Intriguingly, we found that differences in expression and role in inflammation were also observed in patient datasets stratified according to low or high AKR1B1 and AKR1B10 expression. Patients with a high AKR1B10 expression had a more favorable prognosis and GSEA showed that cells from these patients expressed genes that were highly enriched in metabolic processes, as would be expected for an enzyme that has a reductase function. On the other hand, we found that in AKR1B1 overexpressing patients, who had a worse prognosis, the cells showed an enrichment in inflammatory signaling and cell-cell adhesion. This dichotomy is interesting and implies that overexpression of AKR1B1 in CRC may lead to a loss or alteration in its enzymatic function, causing the activation of additional mitogenic signaling processes. This dichotomy is also reflected by a combined gene expression signature where low AKR1B1 and high AKR1B10 expression reflect a better prognostic prediction than the individual genes alone. Future studies are required to establish whether therapeutic targeting of AKR1B1 will yield beneficial effects in CRC.

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## Compliance with ethical standards

**Conflict of interest** None declared.

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